

## Introduction

**Introduction:** Gastroenteritis is a leading cause of death worldwide across all age groups.<sup>1</sup> It is estimated that 1.31 million people died from diarrheal disease in 2015.<sup>1</sup> High-throughput multiplex assays can aid in rapid identification of pathogens that can cause outbreaks of diarrhea and for infection control in healthcare settings. Despite recent introduction of molecular multiplex pathogen detection platforms, there is a limited choice of systems for clinical labs with medium to high-throughput automation.

To address this need, Applied BioCode® has developed an automated high-throughput molecular diagnostic assay system in a 96-well format\*. The BioCode® Gastrointestinal Pathogen Panel (GPP) is an 18-plex molecular assay for detection of gastrointestinal pathogens (Table 1).

**Table 1. Organisms and toxins detected by the BioCode® GPP**

Bacteria	Parasites
<i>Campylobacter</i> spp. ( <i>C. jejuni</i> , <i>C. coli</i> )	<i>Cryptosporidium</i> spp.
<i>Clostridium difficile</i> toxin A/B	<i>Entamoeba histolytica</i>
Enteroaggregative <i>E. coli</i> (EAEC)	<i>Giardia lamblia</i>
Enteropathogenic <i>E. coli</i> (EPEC)	
Enterotoxigenic <i>E. coli</i> (ETEC): LT/ST	
Shiga-toxin producing <i>E. coli</i> (STEC): stx1/stx2	
<i>E. coli</i> O157	<b>Viruses</b>
<i>Shigella</i> spp. /Enteroinvasive <i>E. coli</i> (EIEC)	Adenovirus 40/41
<i>Salmonella</i> spp.	Norovirus GI/GII
<i>Vibrio parahaemolyticus</i>	Rotavirus A
<i>Vibrio</i> spp. (not <i>parahaemolyticus</i> )	
<i>Yersinia enterocolitica</i>	
	<b>RNA Internal Control</b>

## Method

The BioCode® MDx 3000 platform integrates and automates PCR, post-PCR sample handling and detection steps in a 96-well format. Following extraction of nucleic acids from either unpreserved stool or stool in Cary-Blair transport medium, DNA and RNA targets are amplified by one-step RT-PCR. PCR products are captured by target-specific probes coupled to Barcoded Magnetic Beads (BMBs), and the presence of target sequence(s) is detected by a fluorescent conjugate. Qualitative results are determined by a median fluorescent index (MFI) value relative to assay cutoff. For reproducibility, pooled custom NATrol molecular controls (ZeptoMetix) were run in quadruplicate over 5 days.

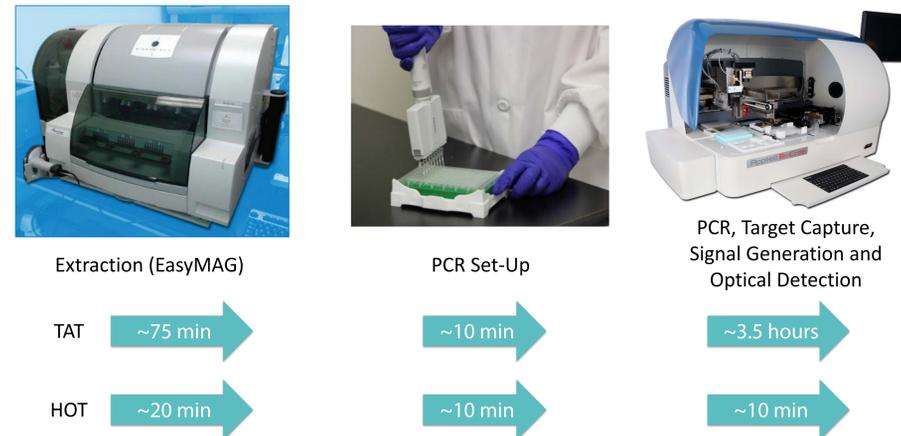
## Reproducibility

**Table 2. Reproducibility of BioCode® GPP on BioCode® MDx 3000.** Reproducibility was evaluated with contrived mixed analytes prepared with NATrol controls from Zeptomatrix. Detection was highly reproducible across 5 days: 99.2% (119/120) overall agreement with inter-assay CVs between 5 - 25%.

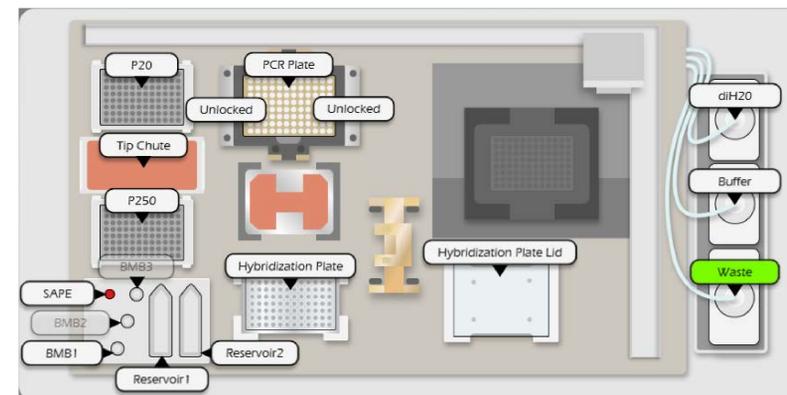
Diarrheagenic <i>E. coli</i>							
		Day 1	Day 2	Day 3	Day 4	Day 5	Between Days
EAEC	MFI	26246	28542	26527	28645	21881	26368
	%CV	3%	4%	5%	4%	6%	10%
EPEC	MFI	28692	30532	30224	32918	26806	29834
	%CV	5%	4%	2%	4%	2%	8%
ETEC (LT)	MFI	5380	7904	5457	7449.3	4938	6225
	%CV	8%	19%	15%	11%	12%	24%
ETEC (ST-a)	MFI	40198	42641	38720	43861	34491	39982
	%CV	3%	2%	1%	2%	6%	9%
ETEC (ST-b)	MFI	23876	30139	25870	28176	20487	25710
	%CV	10%	3%	7%	8%	13%	15%
O157	MFI	29546	29776	29303	30614	22328	28313
	%CV	4%	4%	3%	1%	3%	11%
STEC (stx1)	MFI	7728	8865.3	8998.5	10812	4896	8260
	%CV	9%	5%	12%	3%	11%	25%
STEC (stx2)	MFI	35188	37363	37292	38643	28677	35433
	%CV	4%	5%	3%	5%	5%	11%

Bacteria							
		Day 1	Day 2	Day 3	Day 4	Day 5	Between Days
Campy	MFI	37462	36308	37909	36456	33557	36338
	%CV	1%	2%	2%	2%	3%	5%
C.diff (tcdA)	MFI	11776	14521	14931	16224	11085	13707
	%CV	19%	8%	5%	8%	16%	18%
C. diff (tcdB)	MFI	13417	15297	14929	17694	11679	14603
	%CV	10%	4%	7%	1%	13%	16%
Salmonella	MFI	19166	19800	19401	20588	16399	19071
	%CV	1%	3%	3%	4%	3%	8%
Shigella	MFI	17460	17010	15655	20849	14690	17133
	%CV	9%	27%	7%	3%	11%	18%
V.para	MFI	27404	28503	33279	24705	29440	28666
	%CV	4%	8%	2%	12%	5%	12%
Vibrio spp.	MFI	15008	15408	14494	15442	12612	14593
	%CV	3%	3%	2%	3%	2%	8%
Y.enterocolitica	MFI	27318	30909	25869	31718	21048	27372
	%CV	5%	3%	6%	2%	4%	15%

## BioCode® MDx 3000



**Figure 1. Workflow for BioCode® GI Pathogen Panel.** 192 samples can be processed in an 8-hour shift with minimal hands-on time.



**Figure 2. Schematic for BioCode® MDx 3000 deck layout.** This system incorporates thermal-cycling, BMB-Capture and detection into one automated system. MDx 3000 is designed to run 3 different assays simultaneously in one plate.

## Clinical Performance / Method Comparison

**Table 3. Clinical performance.** 300 unpreserved stool samples were compared to composite results for Luminex xTAG GPP and Sequencing. Overall positive agreement was 96.1% (98/102); negative agreement was 93.4% (183/196). Invalid rate was <1% (2/300).

Target Pathogens	Positives without or with *discordant testing			Resolved agreement (%)	
	BioCode® GPP	Luminex xTAG GPP	Verified Luminex positives*	Negative	Positive
<i>Campylobacter</i> spp.	24	23	22*	99.3	100
<i>Clostridium difficile</i> Toxin A/B	20	17	17	98.9	100
EAEC	10	N/A	N/A	N/A	N/A
EPEC	9	N/A	N/A	N/A	N/A
ETEC	8	4	4	98.6	100
<i>Salmonella</i> spp.	8	15	8*	100	100
<i>Shigella</i> spp./ EIEC	7	7	7	100	100
STEC	11	12	11*	100	100
<i>E. coli</i> O157	1	1	1	100	100
<i>Vibrio parahaemolyticus</i>	1	N/A	N/A	N/A	N/A
<i>Vibrio</i> spp. (not <i>parahaemolyticus</i> )	0	N/A	N/A	N/A	N/A
<i>Yersinia enterocolitica</i>	1	N/A	N/A	N/A	N/A
<i>Cryptosporidium</i> spp.	4	4	4	100	100
<i>Entamoeba histolytica</i>	1	6	1*	100	100
<i>Giardia lamblia</i>	5	26	5*	100	100
Adenovirus (Type 40 & 41)	9	5	5	98.6	100
Norovirus GI/GII	27	26	26	99.6	100
Rotavirus A	9	8	8	99.7	100

**Table 4. Clinical performance.** 100 Cary-Blair samples were compared to composite results for BioFire Filmarray GI Panel and Sequencing. Overall positive agreement was 94.1% (64/68), and negative agreement was 87.5% (28/32). There were no invalid samples.

Target Pathogens	Positive Results without or with *Discordant Testing			Resolved agreement (%)	
	BioCode® GPP	BioFire Filmarray GI Panel	Verified BioFire positives*	Negative	Positive
<i>Campylobacter</i> spp.	8	9	8*	100	100
<i>Clostridium difficile</i> Toxin A/B	13	15	15	100	87
EAEC	7	7	7	100	100
EPEC	11	14	12*	100	92
ETEC	9	9	9	100	100
<i>Salmonella</i> spp.	13	15	14*	100	93
<i>Shigella</i> spp./ EIEC	10	11	10*	100	100
STEC	4	4	4	100	100
<i>E. coli</i> O157	2	N/A	N/A	N/A	N/A
<i>Vibrio parahaemolyticus</i>	0	0	0	100	N/A
<i>Vibrio</i> spp.	0	0	0	100	N/A
<i>Yersinia enterocolitica</i>	1	1	1	100	100
<i>Cryptosporidium</i> spp.	3	3	3	100	100
<i>Entamoeba histolytica</i>	0	0	0	100	N/A
<i>Giardia lamblia</i>	2	2	2	100	100
Adenovirus (Type 40 & 41)	3	2	2	99	100
Norovirus (GI & GII)	8	8	8	99	88
Rotavirus A	8	5	5	97	100

\*Discordant samples tested and resolved by sequencing or repeat testing

## Conclusions

- ❖ The BioCode® Gastrointestinal Pathogen Panel, performed with high clinical accuracy and precision for patient samples and controls.
- ❖ The invalid rate was <1%, resulting in minimal repeats or unresolved samples.
- ❖ Combined with the automated system, the 18-plex molecular panel was intuitive and easy to use.
- ❖ The software guided set up and clear results reports allowed quick implementation with minimal training time.